

Medicinal Plants and their Compounds with Therapeutic Potential in the Treatment of Cancer: an Integrative Review

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Plantas Medicinais e seus Compostos com Potencial Terapêutico no Tratamento do Câncer: Revisão Integrativa

Plantas Medicinales y sus Compuestos con Potencial Terapêutico en el Tratamiento del Câncer: Revisión Integradora

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ABSTRACT

Introduction: The epidemiological and nutritional transitions have been contributing for the exponential growth of cancer. Thus, in order to enhance the activity of antineoplastic drugs, the number of studies using medicinal plants for cancer treatment has increased. **Objective:** To identify medicinal plants and compounds with possible action in the cancer treatment process. **Method:** An integrative review was carried out. For the bibliographic search, the descriptors “neoplasms” and “medicinal plants” and the qualifier “treatment” for the word cancer were used, specifying it. The articles were surveyed in the following databases: Scientific Electronic Library Online (SciELO), Latin American and Caribbean Literature in Health Sciences (LILACS) and Medical Literature Analysis and Retrieval System on-line (MEDLINE) from the PubMed. **Results:** The search and application of the inclusion and exclusion criteria resulted in the selection of 93 articles. The main characteristics of the studies included were *in vitro* assays, with breast cancer cell line and published in English. It was possible to identify six compounds that stood out due to their pro-apoptotic activity in various types of cancer such as curcumin, epicatechin, lupeol, caffeic acid, ursolic acid and berberine. **Conclusion:** The present work offers inputs so that future studies can associate the use of compounds present in medicinal plants with conventional treatment, in order to improve the prognosis of cancer patients. **Key words:** Neoplasms/therapy; Plants, Medicinal/chemistry; Therapeutics; Complementary Therapies.

RESUMO

Introdução: As transições epidemiológica e nutricional têm contribuído para o crescimento exponencial do câncer. Assim, com o intuito de potencializar a atividade das drogas antineoplásicas, aumentou o número de estudos utilizando plantas medicinais para tratamento do câncer. **Objetivo:** Identificar as plantas medicinais e os compostos com possível ação no processo de tratamento do câncer. **Método:** Foi realizada uma revisão integrativa. Para a busca bibliográfica, foram utilizados os descritores “neoplasias” e “plantas medicinais” e o qualificador “tratamento” para a palavra câncer, especificando-a. O levantamento dos artigos foi feito nas seguintes bases de dados: *Scientific Electronic Library OnLine* (SciELO), Literatura Latino-Americana e do Caribe em Ciências da Saúde (LILACS) e *Medical Literature Analysis and Retrieval System Online* (MEDLINE) via PubMed. **Resultados:** A busca e a aplicação dos critérios de inclusão e exclusão resultaram na seleção de 93 artigos. As principais características dos estudos incluídos eram ensaios *in vitro*, com a linhagem de células de câncer de mama e publicados em inglês. Foi possível identificar seis compostos que se destacaram em razão da sua atividade pró-apoptótica em vários tipos de câncer, a exemplo da curcumina, da epicatequina, do lupeol, do ácido cafeico, do ácido ursólico e da berberina. **Conclusão:** O presente trabalho oferece subsídios para que pesquisas futuras possam associar a utilização de compostos presentes em plantas medicinais ao tratamento convencional, com o intuito de melhorar o prognóstico de pacientes oncológicos. **Palavras-chave:** Neoplasias/terapia; Plantas Medicinais/química; Terapêutica; Terapias Complementares.

RESUMEN

Introducción: Las transiciones epidemiológica y nutricional han contribuido al crecimiento exponencial del cáncer. Por lo tanto, para mejorar la actividad de los fármacos antineoplásicos utilizados para tratar la enfermedad, ha aumentado el número de estudios que utilizan plantas medicinales para este propósito. **Objetivo:** Identificar plantas medicinales y compuestos con posible acción en el proceso de tratamiento del cáncer. **Método:** Se realizó una revisión integradora, utilizando los términos de búsqueda validados y utilizados: “neoplasias” y “plantas medicinales”. El calificador “tratamiento” se usó para la palabra cáncer, especificándolo. Los artículos fueron encuestados en las siguientes bases de datos: Biblioteca Científica Electrónica en Línea (SciELO), Literatura Latinoamericana y Caribeña en Ciencias de la Salud (LILACS) y Sistema de Análisis y Recuperación de Literatura Médica en Línea (MEDLINE) vía PubMed. **Resultados:** La búsqueda y la aplicación de los criterios de inclusión y exclusión dieron como resultado la selección de 93 artículos. Las características principales de los estudios incluídos fueron ensayos *in vitro*, con linaje celular de cáncer de mama y publicados en inglés. En relación con los estudios, fue posible identificar seis compuestos que se destacaron por su actividad contra diferentes tipos de cáncer: curcumina, epicatequina, lupeol, ácido cafeico, ácido ursólico y berberina, induciendo la apoptosis por varios mecanismos. **Conclusión:** El presente trabajo, nos permite ofrecer subsidio para que la investigación futura pueda asociar el uso de compuestos presentes en plantas medicinales con el tratamiento convencional, para mejorar el pronóstico de los pacientes con cáncer. **Palabras clave:** Neoplasias/terapia; Plantas Medicinales/química; Terapêutica; Terapias Complementarias.

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INTRODUCTION

Cancer is one of the most prevalent diseases worldwide and the number of harms and deaths related to this condition is growing exponentially¹. Studies attempting to identify the substances for the treatment that can improve patients' survival and quality of life have increased in the last years².

In this context, medicinal plants and functional food can work as potential adjuvant in cancer therapy considering the biological properties of its bioactive compounds³. In addition, the medicinal plants components and functional food have low cost and are easily accessed, yet safe dosages need to be determined⁴.

This study is justified in order to elucidate the use of natural compounds in cancer therapy and stimulate the development of other studies on the theme. The objective of this study was to identify the medicinal plants and compounds with possible pharmacologic action in the process of cancer treatment.

METHOD

Integrative review proposed by methodological reference⁵ consisting of six phases: 1) definition of the research question; 2) search in the databases; 3) data collection; 4) critical analysis of the studies included; 5) discussion of the results; and 6) presentation of the data with unambiguous notification of the results encountered.

The hypothesis to be tested in this study adopted the PICO⁶ strategy where “P” stands for population, patient, or problem, “I”, interest and “Co”, context of the study. Thus, the following research question was elaborated: “What are the medicinal plants and compounds most utilized in cancer treatment?”

The databases Scientific Electronic Library Online (SciELO), Latin America and Caribbean Health Sciences Literature (LILACS) and Medical Literature Analysis and Retrieval System Online (MEDLINE) via PubMed were searched to find the articles.

The criteria to select the articles comprehended all the categories with abstracts and full texts available for analysis and published in Portuguese, English, and Spanish between 2015 and 2019. In addition, the articles selected should respond to the research question.

The following descriptors acknowledged by the Descriptors of Sciences of Health (DeCS) and Medical Subject Headings (MeSH) adopted in the search were: “neoplasms” and “medicinal plants”. In addition, the qualifier “treatment” was used for the descriptor “neoplasm”, for better definition of the aspects. The

Boolean operator “AND” was used to associate the descriptors in the databases.

This study adopted the classification of the articles included per levels of evidences proposed in the literature⁷.

The construction of the database was based in the tool “Instrument of data collecting of the integrative review” proposed by the authors⁸, containing variables of identification of the study as well as the main results and conclusion.

RESULTS

The initial search with the descriptors encountered 22,908 articles and of these, only 172 met the inclusion criteria and responded to the research question. After refining the studies found, 77 were duplicate, in different bases and databases – with the same search tool – which reduced the number to 93 articles that formed the sample.

The results showed that the biggest number of publications were concentrated in 2017 (n=23, 24.7%) and 2018 (n=24, 25.8%). All the articles were in English (n=93, 100%). Of the types of cancer most investigated, breast cancer was quoted in 46.2% (n=43) of the articles, colon cancer in 23.6% (n=22), prostate in 20.4% (n=19), liver, in 19.3% (n=18), kidney cancer in 14% (n=13) and lung cancer in 12.9% (n= 12); in most of the articles, different studies addressed more than one cancer simultaneously.

The type of trial most adopted in the texts selected was *in vitro* quoted in 68.8% (n=64) of the articles, followed by review (integrative and narrative) in 18.3% (n=17). *In vivo* trials were developed in 7.5% (n=7) of the sample and 4.3% (n=4) presented *in vitro* and *in vivo* trials concomitantly. Only one study (1.1%) addressed oncologic patients (double-blind randomized clinical trial).

Regarding evidence, 80.6 % (n=75) of the articles were at level III, 18.3% (n=17) at level II and only one article (1.1%) at level I of the reference adopted.

The main botanic families quoted in the articles were Asteraceae 10.5% (n=10), Fabaceae 8.4% (n=8), Zingiberaceae 7.4% (n=7) and Theaceae 6.3% (n=6). Families as Solanaceae, Betulaceae, Euphorbiaceae, Berberidaceae, Araliaceae, Rutaceae, Malvaceae, Piperaceae, Acanthaceae, Lamiaceae, Phyllanthaceae and Acanthaceae were quoted in at least two articles of the sample.

The results of this study are presented in Tables 1 to 5 describing the plants most investigated with possible effects in cancer treatment with the respective botanic families and active principles encountered.

DISCUSSION

The studies' results concluded that the medicinal plants utilized in the treatment of different types of cancer

included species and classification of many countries and that during the last years, much emphasis has been given to this kind of study. When Fabaceae, Zingiberaceae and Theaceae stand out, it is possible to notice that the

Table 1. Medicinal plants investigated in cancer treatment. Alfenas, MG, 2020

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
1	Azadmehr et al., 2015 ⁹	Breast	Scrophulariaceae	<i>Scrophularia variegata</i>	Unidentified
2	Caamal-Fuentes et al., 2015 ¹⁰	Liver and lung	Fabaceae	<i>Aeschynomene fascicularis</i>	Spinochalcone A
3	Sawadogo et al., 2015 ¹¹	Leukemia	Verbenaceae	<i>Lantana ukambensis</i> (cambará-de-jardim)	Polymethoxyflavones
4	Esmailbeig et al., 2015 ¹²	Leukemia	Lamiaceae	<i>Satureja hortensis</i> (summer savory)	Unidentified
5	Formagio et al., 2015 ¹³	Breast, ovary, lung, and leukemia	Annonaceae	Annonaceae	Unidentified.
6	Ghorbani e Hosseini, 2015 ¹⁴	Breast, colon, and liver	Zingiberaceae	<i>Curcuma longa</i> (saffron)	Curcumin
7	Hosseini et al., 2015 ¹	Liver	Iridaceae	<i>Crocus sativus</i> (true saffron)	Unidentified
8	Jafri et al., 2016 ¹⁵	Breast and cervical cancer	Araliaceae	<i>Hedera nepalensis</i> (ivy)	Lupeol
9	Kuete et al., 2015 ¹⁶	Breast, colon, and prostate	Unidentified	<i>Nauclea pobeguini</i>	Unidentified
10	Kuete e Efferth, 2015 ¹⁷	Kidney and liver	Piperaceae	<i>Piper Capense</i>	Phenolic components
11	Manosroi et al., 2015 ¹⁸	Liver	Unidentified	<i>Ventilago denticulata</i>	Unidentified
12	Mohammed et al., 2015 ¹⁹	Leukemia	Acanthaceae	<i>Andrographis lineata</i>	Flavones
13	Nourazarian et al., 2015 ²⁰	Colon	Fabaceae	<i>Glycyrrhiza glabra</i> (licorice)	Unidentified
14	Rabe et al., 2015 ²¹	Melanoma	Asteraceae	<i>Artemisia khorassanica</i>	Sesquiterpene lactone
15	Sung et al., 2015 ²²	Stomach, breast, prostate	Lamiaceae	<i>Salvia miltiorrhiza</i> (sage)	Dihydrotanshinone I
16	Wen et al., 2015 ²³	Prostate	Apiaceae	<i>Cnidium monnieri</i>	Coumarin
17	Yaacob et al., 2015 ²⁴	Breast	Acanthaceae	<i>Strobilanthes crispus</i>	Lutein
18	Yadav et al., 2015 ²⁵	Breast	Fabaceae	<i>Saraca indica</i> (asoka tree)	Unidentified
19	Ali et al., 2016 ²⁶	Breast	Unidentified	<i>Ochradenus arabicus</i>	Unidentified

Caption: *Compound found in unspecified medicinal plants.

Table 2. Medicinal plants investigated in cancer treatment. Alfenas, MG, 2020

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
20	Asadi-Samani et al., 2016 ²	Breast and prostate	Theaceae	<i>Camellia sinensis</i> (Indian tea)	Epicatechin
21	Chen et al., 2016 ²⁷	Stomach and colon	Rhamnaceae	<i>Rhamnus davurica</i>	Unidentified
22	Du et al., 2016 ²⁸	Colorectal	Thymelaeaceae	<i>Daphne genkwa</i>	Flavonoids
23	Narayanan e Antonisamy, 2016 ²⁹	Breast	Cyatheaceae	<i>Cyathea spp</i> (tree fern)	Unidentified
24	Kim et al., 2016 ³⁰	Colon	Brassicaceae, Apiaceae e Betulaceae	<i>Descurainia sophia</i> , <i>Peucedanum praeruptorum</i> (herb-sophia)	Unidentified
25	Kuete et al., 2016 ³¹	Breast, colon, and prostate	Annonaceae e Passifloraceae	<i>Annona muricata</i> e <i>Passiflora edulis</i> (soursop and passion fruit)	Eugenol and quinones
26	Leelawat e Leelawat, 2016 ³²	Colangio carcinoma	Phyllanthaceae	<i>Phyllanthus emblica</i> (Indian gooseberry)	Unidentified
27	Motawi et al., 2016 ³³	Breast	Unidentified	Unspecified*	Caffeic acid
28	Prasad et al., 2016 ³⁴	Lung, breast and gall bladder	Solanaceae, Phyllanthaceae,	<i>Withani somnifera</i> , <i>Phyllanthus amarus</i> (Indian ginseng and leaf flower)	Unidentified
29	Pereira et al., 2016 ³⁵	Leukemia	Geraniaceae	<i>Pelargonium sidoides</i>	Fluorouracil
30	Sapio et al., 2017 ³⁶	Stomach and mouth	Labiatae	<i>Coleus forskohlii</i> (boldo-brasileiro)	Forskolin
31	Subramani et al., 2016 ³⁷	Pancreas	Meliaceae	<i>Azadirachta indica</i> (Amargosa)	Nimbolide
32	Sun et al., 2016 ³⁸	Ovary	Rosaceae	<i>Pyracantha fortuneana</i> (Piracanta)	Polysaccharides
33	Thomas et al., 2016 ³⁹	Leukemia, breast, and kidney	Asteraceae	<i>Vernonia condensata</i> (boldo-baiano)	Vernolide A
34	Teoh et al., 2016 ⁴⁰	Breast	Acanthaceae	<i>Clinacanthus nutans</i>	Unidentified
35	Uche et al., 2016 ⁴¹	Ovary	Menispermaceae	<i>Triclisia subcordata</i>	Cicleanine
36	Zhao et al., 2016 ⁴²	Breast	Ranunculacea, Berberidaceae	<i>Hydrastis canadensis</i> , <i>Berberis aristata</i> (Balisse fruit)	Berberine
37	Attar et al., 2017 ⁴³	Ovary	Phyllanthaceae	<i>Phyllanthus spp</i>	Corilagin
38	Bhandari et al., 2017 ⁴⁴	Prostate, breast, and cervical cancer	Alliaceae	<i>Allium wallichii</i> (garlic)	Terpenoids and flavonoids

Caption: *Compound found in unspecified medicinal plants.

Table 3. Medicinal plants investigated in cancer treatment. Alfenas, MG, 2020

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
39	Chandel et al., 2017 ⁴⁵	Uterus and colon	Rubiaceae	<i>Anthocephalus cadamba</i>	Alkaloids
40	Rodrigues et al., 2017 ⁴⁶	Prostate and melanoma	Combretaceae	<i>Terminalia fagifolia</i> (Caatinga-de-porco)	Unidentified
41	Dushimemaria et al., 2017 ⁴⁷	Breast, melanoma, and kidney	Fabaceae	<i>Guibourtia coleosperma</i>	Unidentified
42	Harsha Raj et al., 2017 ⁴⁸	Breast	Myrtaceae, Musaceae	<i>Eugenia jambolana</i> , <i>Musa Paradisiaca</i> . (Indian blackberry and banana tree)	Unidentified
43	Kooti et al., 2017 ⁴⁹	Breast, colon, and liver	Theaceae, Amaryllidace, Zingiberaceae	<i>Camellia sinensis</i> , <i>Allium sativum</i> , <i>Curcuma longa</i> (Indian tee, garlic, saffron)	Epicatechin, allicin and curcumin
44	Kuete et al., 2017 ⁵⁰	Lung, liver and breast	Asteraceae	<i>Elephantopus mollis</i> (tobacco weed, soft elephantsfoot erva-moli)	Flavonoids and polyphenols
45	Mbaveng et al., 2017 ⁵¹	Breast, prostate, colon	Piperaceae	<i>Piper capins</i>	Unidentified
46	Liang et al., 2017 ⁵²	Leukemia, liver and uterus	Nyctaginaceae, Brassicaceae	<i>Bougainvillea spectabilis</i> , <i>Brassica campestris</i> (três-marias turnip, wormseed)	Unidentified
47	Mbele et al., 2017 ⁵³	Prostate, kidney, and gall bladder	Zingiberaceae	<i>Zingiber officinale</i> (ginger)	Unidentified
48	Nguyen et al., 2017 ⁵⁴	Pancreas, colon, ovary, lung, and skin	Rutaceae	<i>Paramignya trimera</i>	Ostruthin
49	Okubo et al., 2017 ⁵⁵	Leukemia	Ranunculaceae and Rutaceae	<i>Coptis japonica</i> e <i>Phellodendron amurense</i>	Berberine
50	Pandey, 2017 ⁵⁶	Melanoma	Fabaceae	<i>Bauhinia variegata</i> (pata-de-vaca)	Flavonoids
51	Rahman et al., 2017 ⁵⁷	Prostate	Boraginaceae	<i>Cordia dichotoma</i>	Flavonoids
52	Rajavel et al., 2017 ⁵⁸	Lung	Malvaceae	<i>Grewia tiliaefolia</i>	Unidentified
53	Roman Junior et al., 2017 ⁵⁹	Breast and lung	Zingiberaceae	<i>Alpinia zerumbet</i> (false cardamom)	5,6-Dehydrokavain

to be continued

Table 3. continuation

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
54	Thakor et al., 2017 ⁶⁰	Kidney	Unidentified	Unspecified*	Maslinic acid
55	Torquato et al., 2017 ⁶¹	Breast, prostate, colon, liver	Zingiberaceae and Theaceae	<i>Curcuma longa</i> e <i>Camellia sinensis</i> (saffron and Indian tea)	Curcumin and Epigallocatechin -3
56	Xia et al., 2017 ⁶²	Breast, colon, and prostate	Lamiaceae	<i>Scutellaria baicalensi</i> (Baikal skullcap)	Baicalin
57	Yadav et al., 2017 ⁶⁴	Breast	Asteraceae	<i>Eclipta alba</i> (erva-botão)	Unidentified
58	Zhang et al., 2017 ⁶³	Breast	Ranunculaceae	<i>Aconitum coreanum</i> (Wolfsbane Potion)	Polysaccharides
59	Zhong et al., 2017 ⁶⁴	Lung, breast, liver, cervical and prostate.	Dryopteridaceae	<i>Dryopteris fragrans</i>	Unidentified
60	Abu-Darwish e Efferth, 2018 ⁶⁵	Breast, prostate and kidney	Solanaceae	<i>Withania somnifera</i> (Indian ginseng)	Unidentified

Caption: *Compound found in unspecified medicinal plants.

Table 4. Medicinal plants investigated in cancer treatment. Alfenas, MG, 2020

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
61	Al-Dabbagh et al., 2018 ⁶⁶	Liver	Apocynaceae	<i>Rhazya stricta</i>	Flavonoids
62	Asadi-Samani et al., 2018 ⁶⁷	Prostate	Euphorbiaceae and Asteraceae	<i>Euphorbia szovitsii</i> e <i>Achillea wilhelmsii</i>	Unidentified
63	Bonam et al., 2018 ⁶⁸	Breast, kidney, liver and colon	Zingiberaceae	<i>Curcuma longa</i> e <i>curcuma zedoaria</i> (saffron and zedoária)	Curcumin
64	Chen et al., 2018 ⁶⁹	Uterus, melanoma	Berberidaceae	Gênero <i>Epimedium</i>	Icaritin
65	Dong et al., 2018 ⁷⁰	Pancreas	Fabaceae	<i>Pao Pereira</i> (pau-pereira)	Unidentified
66	Escher et al., 2018 ⁷¹	Colon, liver and lung	Asteraceae	<i>Centaurea cyanus</i> (escovinha)	Chlorogenic, caffeic and ferulic acids
67	Esghaei et al., 2018 ⁷²	Colon	Theaceae	<i>Camellia sinensis</i> (Indian tea)	Epicatechin
68	Fort et al., 2018 ⁷³	Prostate	Euphorbiaceae	<i>Cnidoscylus chayamansa</i>	Unidentified
69	Gomes et al., 2018 ⁷⁴	Breast, kidney and prostate	Asteraceae	<i>Solidago chilensis</i> (Brazilian arnica)	Diterpene solidagenone

to be continued

Table 4. continuation

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
70	Hong et al., 2018 ⁷⁵	Breast	Unidentified	Unspecified*	Apigenin and luteolin
71	Kuete et al., 2018 ⁷⁶	Breast, liver and colon	Unidentified	<i>Ptycholobium contortum</i>	β -amirine and lupeol
72	Maciel et al., 2018 ⁷⁷	Liver, colorectal and lung	Malvaceae	<i>Hibiscus sabdariffa</i>	Anthocyanins and flavonoids
73	Malvicini et al., 2018 ⁷⁸	Colon	Poaceae	<i>Deschampsia antarctica</i> (Antarctic hair grass)	Antartina
74	Ogunlaja et al., 2018 ⁷⁹	Breast and colon	Moraceae	<i>Ficus burtt-davyi</i>	Catechin and lupeol
75	Saeed et al., 2018 ⁸⁰	Breast, colon, and liver	Betulaceae	<i>Betula pendula</i> (White birch)	Betulinic acid
76	Santos et al., 2018 ⁸¹	Liver, colorectal	Fabaceae, Theaceae Aquifoliaceae	<i>Aspalathus linearis</i> , <i>Camellia sinensis</i> e <i>Ilex paraguariensis</i>	Kaempferol, catechin, epicatechin
77	Sethi et al., 2018 ⁸²	Breast, kidney, and colon	Fabaceae	<i>Trigonella foenum-graecum</i> (feno-grego)	Diosgenin
78	Sodrul et al., 2018 ⁸³	Breast, kidney and colon	Araliaceae	<i>Panax ginseng</i> (ginseng)	Ginsenosides
79	Soyingbe et al., 2018 ⁸⁴	Breast, uterus and colon	Apiaceae, Canellaceae e Curtisiaceae	<i>Centella asiatica</i> , <i>Warburgia salutaris</i> e <i>Curtisia dentata</i>	Unidentified

Caption: *Compound found in unspecified medicinal plants.

Table 5. Medicinal plants investigated in cancer treatment. Alfenas, MG, 2020

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
80	Nguyen et al., 2019 ³	Pancreas	-	<i>Paramignya trimera</i>	Gallic acid and caffeic acid
81	Tayeh e Ofir, 2018 ⁸⁵	Lymph nodes	Asteraceae	<i>Asteriscus graveolens</i> (rue)	Unidentified
82	Uen et al., 2018 ⁸⁶	Mouth	Solanaceae	<i>Solanum nigrum</i> (nightshades)	Unidentified
83	Zhang et al., 2018 ⁸⁷	Uterus	Asteraceae	<i>Dryopteris fragrans</i>	Unidentified
84	Asl et al., 2018 ⁸⁸	Stomach	Linaceae	<i>Linum álbum</i> (fedegoso)	Unidentified
85	Aumeeruddy e Mahomoodally, 2019 ⁸⁹	Breast	Piperaceae, Piperaceae and Ranunculaceae	<i>Piper nigrum</i> , <i>Piper longum</i> e <i>Nigella sativa</i> (black pepper)	Piperine, Sulforaphane and thymochinone
86	Cordeiro et al., 2019 ⁹⁰	Lung	Lythraceae	<i>Lafoensia pacari</i> (pacari)	Procyanidin

to be continued

Table 5. continuation

Identification of the article	Author/year	Types of cancer	Botanic family	Medicinal plant (popular name)	Active principle
87	De et al., 2019 ⁹¹	Lung and cervical cancer	Dilleniaceae	<i>Dillenia pentagyn</i> (elephant apple)	Unidentified
88	Iqbal et al., 2019 ⁹²	Skin	Zingiberaceae	<i>Curcuma longa</i> (saffron)	Curcumin
89	Lim et al., 2019 ⁹³	Colon, breast cancer and leukemia	Amaryllidaceae	<i>Crinum amabile</i> (açucena-do-brejo)	Unidentified
90	Mfengwana et al., 2019 ⁹⁴	Prostate, breast	Asparagaceae and Asteraceae	<i>Asparagus laricinus e Senecio asperulus</i> (aspargus)	Unidentified
91	Sinha et al., 2019 ⁹⁵	Kidney	Unidentified	Unspecified*	Lupeol
92	Wang et al., 2019 ⁹⁶	Leukemia	Meliaceae	<i>Melia azedarach</i> (cinnamon)	Unidentified
93	Zahra et al., 2019 ⁹⁷	Breast, liver and larynx	Zingiberaceae	<i>Alpinia zerumbet</i> (false cardamom)	Unidentified

Caption: *Compound found in unspecified medicinal plants.

substances encountered in the plants of these families may lead to remarkable findings for cancer treatment with important antiproliferative effects both *in vitro* and *in vivo*. Therefore, this review attempted to guide future studies with other species of these families in order to detail the mechanisms of action and application in cancer⁴ treatment.

The type of cancer most described in this survey was breast cancer, associated with extracts of medicinal plants in cells cultures mainly. According to some authors^{15,70}, breast cancer is one of the main causes of death in women worldwide and the use of alternative therapies with natural products may increase the survival of these patients effectively and safely.

Some investigators reported the necessity of identifying the compounds found in medicinal plants responsible for the antineoplastic effect⁶⁵. In this review, it was possible to evaluate that the compounds curcumin, epicatechin, lupeol, caffeic acid, ursolic acid and berberine stood out because of their activity against several cancer types, especially in *in vitro* studies where these compounds were isolated from medicinal plants used in assays. Curcumin can be encountered in *Curcuma longa* mainly and demonstrated effect on the reduction of the cellular viability of breast and colon cancer by promoting accumulation of reactive oxygen species (ROS), which can promote cellular stress and trigger the process of apoptosis^{32,81}.

Another study described that curcumin is a potent inhibitor of NF- κ B (nuclear factor kappa B), and anticancer agent since this protein complex (NF- κ B) is

related to the response to stress stimuli and its deregulation is connected to the development of tumors^{12,76}. In addition, curcumin was associated with the cell cycle arrest in phase G2/S, inducing apoptosis. These findings associated with easy access and low cost of the plant may indicate the possibility of application of curcumin in the therapy of prostate, breast, and colon tumors, potentializing the effect of conventional therapy⁸¹. It is important to highlight that most part of the studies analyzing the action of this component was carried out with *in vitro* and *in vivo* assays and although these assays show the concentration of the extract utilized, it is difficult to extrapolate to human beings in what concerns the determination of the dosage; more clinical trials and study of mathematical models that potentially may determine the posology to be used to reach the expected results are necessary¹.

Epicatechin, a flavonoid that can be found in plants as *Camellia sinensis*, known as India tea, green tea, white tea, red tea among other popular names was evaluated for inhibiting the growth of cancer cells with low toxicity for healthy cells^{2,61}. The mechanism of action of this substance was associated with the increase of the concentration of caspases, that can be defined as essential and constitutive proteases of the apoptosis process.

The alternative treatment with medicinal plants was able to increase the concentration of initiator caspases (as numbers 8 and 9), whose function is to cleavage the inactive preformed caspases and the concentration of effector caspases (as number 3, for instance), that cleavage other substrates, triggering the apoptotic cascade⁴⁶.

This characteristic allows the use of the compound in anticancer therapy because it is safe for normal cells and exerts antiproliferative effect in tumor tissues³³.

Lupeol of the class of pentacyclic triterpenes encountered in common fruits as olive, mango and fig showed promising effects in cancer treatment⁷². Studies^{41,42} demonstrated that this compound was able to develop several functions, among them, inhibition of proliferation, migration and invasion of kidney carcinoma and gallbladder, further to inducing cells to apoptosis.

Possibly, lupeol-induced cellular death mechanism is related with the suppression of matrix metalloproteinases (MMP-9), an enzyme that degrades the basal membrane, allowing that factors involved with the carcinogenic process invade the blood flow, giving origin to metastases. This compound can be encountered in the plant of the species *Hedera nepalensis* and the ethanolic extract was able to inhibit the growth in more than 60% of cells culture of breast and cervical cancer, indicating possible application in oncologic therapy⁸⁷.

Similarly, other studies demonstrated that lupeol has cytotoxic activity for lineages of breast, liver and colon cancer and can be a promising candidate as drug for therapy against cancer because of the modulation of the mitochondrial dynamic influencing processes that interfere in the integrity of the membrane, provoking apoptosis. However, posology evaluation will be necessary in future studies for application in human beings as the articles address *in vitro* and *in vivo* trials^{28,84}.

Another compound identified in the studies of this review was caffeic acid, a phenolic acid whose property is its binding to other compounds with important antiproliferative and antioxidant activity. This acid activated the process of apoptosis in breast cancer cell through modulation of caspases and fragmentation of DNA, destroying tumor cells⁷².

In addition, it was associated with the control of the cellular cycle related to the genes of the family Bcl-2 and to the proteins originated from said genes. These compounds belong to the family of antiapoptotic proteins (Bcl2, for example) as well as by proapoptotic proteins (like Bax), the compound was able to regulate Bcl-2 negatively and reduce the relation Bcl2/Bax⁴⁵. Therefore, the application of the caffeic acid as adjuvant in cancer treatment can be viable since this compound also demonstrated low cytotoxicity to normal cells and can preserve healthy tissues in patients with cancer³.

The ursolic acid, a pentacyclic triterpenoid identified in the epicuticular waxes of the apples and widely found in fruits peels and medicinal plants was able to induce the apoptosis of skin cancer cells by increasing the concentration of caspases in special effector caspases

number 3 responsible for substrate cleavage, promoting sequencing that leads to cellular death⁷⁶. The results encountered showed dose-dependent efficacy, which suggests that the adjuvant application in cancer treatment should be done with the compound isolated in order to potentialize the effects of the conventional therapy⁹¹.

Finally, the compound berberine, a natural benzyloquinoline alkaloid extracted from rhizomes and roots of several plants, as the species *Berberis aristata*, evaluated as opposed to breast cells cancer. This compound was effective in preparing the cells to cisplatin (antineoplastic) with activation of pro-apoptotic factors also related to the increase of caspases, specially caspase 9 responsible for the initial phases of the apoptosis process⁶⁶. It can be indicated as adjuvant therapy to the clinical treatment, mainly in patients with cancer who suffered metastasis because of its potentializing activity of the effects of antimetastatic drugs⁴⁹.

Considering the findings of the present integrative review, studies of *in vitro* and *in vivo* dosages are relevant and necessary to support its potential application in human beings according to a study where four patients with liver cancer who used 100 mg/day of encapsulated dry saffron (*Crocus sativus*) were compared with the placebo group, associated with chemotherapy. The observations showed that in the group treated with saffron, two patients responded partially or completely to the elective treatment and in the placebo group, no response was observed, which can suggest a possible application of these compounds in the protocol of treatment of patients with malignant neoplasms¹.

The discoveries and information presented in the studies selected and included in the analysis of this review can bring elements for new studies related to the identification of active principles of the medicinal plants as adjuvant therapy in human beings in order to potentialize the effects of the conventional therapy to prepare the cells to absorb the drugs utilized and increase the rate of cancer cellular death, leading to better prognosis and reversing the course of the disease.

CONCLUSION

The results of the studies included in this review allowed to conclude that the substances most investigated in the medicinal plants for cancer treatment are curcumin, epicatechin, lupeol, caffeic acid, ursolic acid and berberine. These compounds have been evaluated mostly in *in vitro* and *in vivo* assays due to their antineoplastic potential and specific effects as cellular antiproliferation, increase of pro-apoptotic proteins and reduction of factors that lead tumor cells to metastasize.

With this, the studies provide elements for future studies to associate the use of the compounds found in the medicinal plants to the conventional treatment (radio or chemotherapy) in order to potentialize the effects of this treatment and improve the prognosis of oncologic patients.

Future studies with different methods must be carried out to define better the mechanisms of action through which these components act, safe doses for human beings and potential adverse effects.

CONTRIBUTIONS

The authors contributed equally for the study conception and/or design, collection, analysis and interpretation of the data, wording, critical review and approved the final version to be published.

DECLARATION OF CONFLICT OF INTERESTS

There is no conflict of interests to declare.

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REFERENCES

- Hosseini A, Mousavi SH, Ghanbari A, et al. Effect of saffron on liver metastases in patients suffering from cancers with liver metastases: a randomized, double blind, placebo-controlled clinical trial. *Avicenna J Phytomed.* 2015;5(5):434-40. doi: <http://doi.org/10.22038/ajp.2015.4667>.
- Asadi-Samani M, Kooti W, Aslani E, et al. A systematic review of Iran's medicinal plants with anticancer effects. *J Evid Based Complementary Altern Med.* 2016;21(2):143-53. doi: <http://doi.org/10.1177/2156587215600873>.
- Nguyen VT, Scarlett CJ. Cytotoxic activity of extracts and fractions from *Paramignya trimera* root and *Phyllanthus amarus* against pancreatic cancer cell lines. *J Cancer Res Ther.* 2019;15(1):245-9. doi: http://doi.org/10.4103/jcrt.JCRT_85_18.
- Yadav NK, Arya RK, Dev K, et al. Alcoholic extract of *eclipta alba* shows in vitro antioxidant and anticancer activity without exhibiting toxicological effects. *Oxid Med Cell Longev.* 2017;2017:9094641. doi: <http://doi.org/10.1155/2017/9094641>.
- Souza MT, Silva MD, Carvalho R. Revisão integrativa: o que é e como fazer. *Einstein (São Paulo).* 2010;8(1):102-6. doi: <http://doi.org/10.1590/s1679-45082010rw1134>.
- Sousa CKL, Silva ACO, Silva AL, et al. As evidências científicas da bronquiectasia: etiologia, diagnóstico e formas de tratamento. *Braz J Surg Clin Res [Internet].* 2019 [acesso 2019 set 18];26(3):78-83. Available from: https://www.mastereditora.com.br/periodico/20190520_090939.pdf.
- Melnyk BM, Fineout-Overholt E, Feinstein NF, et al. Nurses' perceived knowledge, beliefs, skills, and needs regarding evidence-based practice: implications for accelerating the paradigm shift. *Worldviews Evid Based Nurs.* 2004;1(3):185-93. doi: <http://doi.org/10.1111/j.1524-475X.2004.04024.x>.
- Fernandes DL. Representações sociais de adolescentes sobre o câncer de colo do útero [dissertação na Internet]. Recife: Universidade Federal de Pernambuco; 2011. [acesso 2019 ago 20]. 87 p. Available from: <https://repositorio.ufpe.br/handle/123456789/9254>.
- Azadmehr A, Hajiaghae R, Baradaran B, et al. Apoptosis cell death effect of *scrophularia variegata* on breast cancer cells via mitochondrial intrinsic pathway. *Adv Pharm Bull.* 2015;5(3):443-6. doi: <http://doi.org/10.15171/apb.2015.060>.
- Caamal-Fuentes EE, Peraza-Sánchez SR, Torres-Tapia LW, et al. Isolation and identification of cytotoxic compounds from *aeschynomene fascicularis*, a mayan medicinal plant. *Molecules.* 2015;20(8):13563-74. doi: <http://doi.org/10.3390/molecules200813563>.
- Sawadogo WR, Cerella C, Al-Mourabit A, et al. Cytotoxic, antiproliferative and pro-apoptotic effects of 5-Hydroxyl-6,7,3',4',5'-Pentamethoxyflavone Isolated from *Lantana ukambensis*. *Nutrients.* 2015;7(12):10388-97. doi: <http://doi.org/10.3390/nu7125537>.
- Esmailbeig M, Kouhpayeh SA, Amirghofran Z. An investigation of the growth inhibitory capacity of several medicinal plants from Iran on tumor cell lines. *Iran J Cancer Prev.* 2015;8(5):e4032. doi: <http://doi.org/10.17795/ijcp-4032>.
- Formagio ASN, Vieira MC, Volobuff CRF, et al. In vitro biological screening of the anticholinesterase and antiproliferative activities of medicinal plants belonging to Annonaceae. *Braz J Med Biol Res.* 2015;48(4):308-15. doi: <http://doi.org/10.1002/ptr.5660>.
- Ghorbani A, Hosseini A. Cancer therapy with phytochemicals: evidence from clinical studies. *Avicenna J Phytomed.* 2015;5(2):84-97. doi: <http://doi.org/10.22038/ajp.2015.3872>.
- Jafri L, Saleem S, Kondrytuk TP, et al. *Hedera nepalensis* K. Koch: a novel source of natural cancer chemopreventive and anticancerous compounds. *Phytother Res.* 2016;30(3):447-53. doi: <https://doi.org/10.1002/ptr.5546>.
- Kuete V, Sandjo LP, Mbaveng AT, et al. Cytotoxicity of selected cameroonian medicinal plants and *Nauclea pobeguini* towards multi-factorial drug-resistant cancer cells. *BMC Complement Altern Med.* 2015;15:309. doi: <http://doi.org/10.1186/s12906-015-0841-y>.

17. Kuete V, Efferth T. African flora has the potential to fight multidrug resistance of cancer. *Biomed Res Int.* 2015;2015:914813. doi: <http://doi.org/10.1155/2015/914813>.
18. Manosroi A, Akazawa H, Kitdamrongtham W, et al. Potent antiproliferative effect on liver cancer of medicinal plants selected from the Thai/Lanna medicinal plant recipe database "MANOSROI III." *Evid Based Complement Alternat Med.* 2015;2015:397181. doi: <http://doi.org/10.1155/2015/397181>.
19. Mohammed A, Chiruvella KK, Rao YK, et al. In vitro production of echinoidinin, 7-O-methylogonin from callus cultures of *Andrographis lineata* and their cytotoxicity on cancer cells. *PLoS One.* 2015;10(10):e0141154. doi: <http://doi.org/10.1371/journal.pone.0141154>.
20. Nourazarian SM, Nourazarian A, Majidinia M, et al. Effect of root extracts of medicinal herb *Glycyrrhiza glabra* on HSP90 gene expression and apoptosis in the HT-29 colon cancer cell line. *Asian Pac J Cancer Prev.* 2015;16(18):8563-6. doi: <http://doi.org/10.7314/APJCP.2015.16.18.8563>.
21. Rabe ST, Emani SA, Iranshahi M, et al. Anti-cancer properties of a sesquiterpene lactone-bearing fraction from *Artemisia khorassanica*. *Asian Pac J Cancer Prev.* 2015;16(3):863-8. doi: <http://doi.org/10.7314/APJCP.2015.16.3.863>.
22. Sung B, Chung HS, Kim M, et al. Cytotoxic effects of Solvent-Extracted active components of *Salvia miltiorrhiza* Bunge on human cancer cell lines. *Exp Ther Med.* 2015;9(4):1421-8. doi: <http://doi.org/10.3892/etm.2015.2252>.
23. Wen YC, Lee WJ, Tan P, et al. By inhibiting snail signaling and miR-23a-3p, osthole suppresses the EMT-mediated metastatic ability in prostate cancer. *Oncotarget.* 2015;6(25):21120-36. doi: <http://doi.org/10.18632/oncotarget.4229>.
24. Yaacob NS, Yankuzo HM, Devaraj S, et al. Anti-tumor action, clinical biochemistry profile and phytochemical constituents of a pharmacologically active fraction of *S. crispus* in NMU-induced rat mammary tumour model. *PLoS One.* 2015;10(5):e0126426. doi: <http://doi.org/10.1371/journal.pone.0126426>.
25. Yadav NK, Saini KS, Hossain Z, et al. *Saraca indica* bark extract shows in vitro antioxidant, antibreast cancer activity and does not exhibit toxicological effects. *Oxid Med Cell Longev.* 2015;2015:205360. doi: <http://doi.org/10.1155/2015/205360>.
26. Ali MA, Farah MA, Al-Hemaid FM, et al. Assessment of biological activity and UPLC-MS based chromatographic profiling of ethanolic extract of *Ochradenus arabicus*. *Saudi J Biol Sci.* 2016;23(2):229-36. doi: <http://doi.org/10.1016/j.sjbs.2015.02.010>.
27. Chen G, Li X, Saleri F, et al. Analysis of flavonoids in *Rhamnus davurica* and its antiproliferative activities. *Molecules.* 2016;21(10):1275. doi: <http://doi.org/10.3390/molecules21101275>.
28. Du WJ, Yang XL, Song ZJ, et al. Antitumor activity of total flavonoids from *Daphne genkwa* in colorectal cancer. *Phytother Res.* 2016;30(2):323-30. doi: <http://doi.org/10.1002/ptr.5540>.
29. Narayanan J, Antonisamy JMA. Ethanol extracts of selected *Cyathea* species decreased cell viability and inhibited growth in MCF 7 cell line cultures. *J Acupunct Meridian Stud.* 2016;9(3):151-5. doi: <http://doi.org/10.1016/j.jams.2016.04.004>.
30. Kim HY, Kim J, Ha Thi HT, et al. Evaluation of anti-tumorigenic activity of BP3B against colon cancer with patient-derived tumor xenograft model. *BMC Complement Altern Med.* 2016;16(1):473. doi: <http://doi.org/10.1186/s12906-016-1447-8>.
31. Kuete V, Dzotam JK, Voukeng IK, et al. Cytotoxicity of methanol extracts of *Annona muricata*, *Passiflora edulis* and nine other Cameroonian medicinal plants towards multi-factorial drug-resistant cancer cell lines. *Springerplus.* 2016;5(1):1666. doi: <http://doi.org/10.1186/s40064-016-3361-4>.
32. Leelawat S, Leelawat K. Molecular mechanisms of cholangiocarcinoma cell inhibition by medicinal plants. *Oncol Lett.* 2016;13(2):961-6. doi: <http://doi.org/10.3892/ol.2016.5488>.
33. Motawi TK, Abdelazim SA, Darwish HA, et al. Modulation of tamoxifen cytotoxicity by caffeic acid phenethyl ester in MCF-7 breast cancer cells. *Oxid Med Cell Longev.* 2016;2016:3017108. doi: <http://doi.org/10.1155/2016/301710>.
34. Prasad NR, Muthusamy G, Shanmugam M, et al. South Asian medicinal compounds as modulators of resistance to chemotherapy and radiotherapy. *Cancers (Basel).* 2016;8(3):32. doi: <http://doi.org/10.3390/cancers8030032>.
35. Pereira A, Bester M, Soundy P, et al. Anti-proliferative properties of commercial *Pelargonium sidoides* tincture, with cell-cycle G0/G1 arrest and apoptosis in Jurkat leukaemia cells. *Pharm Biol.* 2016;54(9):1831-40. doi: <http://doi.org/10.3109/13880209.2015.1129545>.
36. Sapio L, Gallo M, Illiano M, et al. The natural cAMP elevating compound Forskolin in cancer therapy: is it time? *J Cell Physiol.* 2017;232(5):922-7. doi: <http://doi.org/10.1002/jcp.25650>.
37. Subramani R, Gonzalez E, Arumugam A, et al. Nimbolide inhibits pancreatic cancer growth and metastasis through ROS-mediated apoptosis and inhibition of epithelial-to-mesenchymal transition. *Sci Rep.* 2016;6:19819. doi: <http://doi.org/10.1038/srep19819>.
38. Sun Q, Dong M, Wang Z, et al. Selenium-enriched polysaccharides from *Pyracantha fortuneana* (Se-PFPs)

- inhibit the growth and invasive potential of ovarian cancer cells through inhibiting β -catenin signaling. *Oncotarget*. 2016;7(19):28369-83. doi: <http://doi.org/10.18632/oncotarget.8619>.
39. Thomas E, Gopalakrishnan V, Somasagara RR, et al. Extract of *Vernonia condensata*, Inhibits tumor progression and improves survival of tumor-allograft bearing mouse. *Sci Rep*. 2016;6: 23255. doi: <http://doi.org/10.1038/srep23255>.
 40. Teoh PL, Cheng AYF, Liau M, et al. Chemical composition and cytotoxic properties of *Clinacanthus nutans* root extracts. *Pharm Biol*. 2016;55(1):394-401. doi: <http://doi.org/10.1080/13880209.2016.1242145>.
 41. Uche FI, Drijfhout FP, McCullagh J, et al. Cytotoxicity effects and apoptosis induction by bisbenzylisoquinoline alkaloids from *Triclisia subcordata*. *Phyther Res*. 2016;30(9):1533-9. doi: <http://doi.org/10.1002/ptr.5660>.
 42. Zhao Y, Jing Z, Li Y, et al. Berberine in combination with cisplatin suppresses breast cancer cell growth through induction of DNA breaks and caspase-3-dependent apoptosis. *Oncol Rep*. 2016;36(1):567-72. doi: <http://doi.org/10.3892/or.2016.4785>.
 43. Attar R, Cincin ZB, Bireller ES, et al. Apoptotic and genomic effects of corilagin on SKOV3 ovarian cancer cell line. *Onco Targets Ther*. 2017; 10:1941-6. doi: <http://doi.org/10.2147/OTT.S135315>.
 44. Bhandari J, Muhammad BT, Thapa P, et al. Study of phytochemical, anti-microbial, anti-oxidant, and anti-cancer properties of *Allium wallichii*. *BMC Complement Altern Med*. 2017;17:102. doi: <http://doi.org/10.1186/s12906-017-1622-6>.
 45. Chandel M, Kumar M, Sharma U, et al. Investigations on Antioxidant, Antiproliferative and COX-2 Inhibitory Potential of Alkaloids from *Anthocephalus cadamba* (Roxb.) Miq. Leaves. *Chem Biodivers*. 2017;14(4):2-11. doi: <http://doi.org/10.1002/cbdv.20160037>. 10.1002/cbdv.201600376
 46. Rodrigues PSM, Bertolin AO, Fucase TM, et al. Assessment of the cytotoxic activity of ethanol extracts from the bark and leaves of *Terminalia fagifolia* Mart. on normal and tumor cells. *J Health Biol Sci*. 2017;5(1):16-23. doi: <http://doi.org/10.12662/2317-3076jhbs.v5i1.1068.p.16-23.2017>.
 47. Dushimemaria F, Preez CID, Mumbengegwi DR. Randomized anticancer and Cytotoxicity activities of *Guibourtia Coleosperma* and *Diospyros Chamaethamnus*. *Afr J Tradit Complement Altern Med*. 2017;14(4):1-7. doi: <http://doi.org/10.21010/ajtcam.v14i4.1>.
 48. Harsha Raj M, Ghosh D, Banerjee R, et al. Suppression of VEGF-induced angiogenesis and tumor growth by *Eugenia jambolana*, *Musa paradisiaca*, and *Coccinia indica* extracts. *Pharm Biol*. 2017;55(1):1489-99. doi: <http://doi.org/10.1080/13880209.2017.1307422>.
 49. Kooti W, Servatyari K, Behzadifar M, et al. Effective medicinal plant in cancer treatment, part 2: review study. *J Evid Based Complementary Altern Med*. 2017;22(4):982-95. doi: <http://doi.org/10.1177/2156587217696927>.
 50. Kuete V, Fokou FW, Karaosmanoğlu O, et al. Cytotoxicity of the methanol extracts of *Elephantopus mollis*, *Kalanchoe crenata* and 4 other Cameroonian medicinal plants towards human carcinoma cells. *BMC Complement Altern Med*. 2017;17(1):280. doi: <http://doi.org/10.1186/s12906-017-1793-1>.
 51. Mbaveng AT, Kuete V, Efferth T. Potential of central, Eastern and Western Africa medicinal plants for cancer therapy: spotlight on resistant cells and molecular targets. *Front Pharmacol*. 2017;8:343. doi: <http://doi.org/10.3389/fphar.2017.00343>.
 52. Liang C, Pan H, Li H, et al. In vitro anticancer activity and cytotoxicity screening of phytochemical extracts from selected traditional Chinese medicinal plants. *J BUON [Internet]*. 2017 [cited 2019 Aug 29];22(2):543-51. Available from: <https://www.jbuon.com/archive/22-2-543.pdf>.
 53. Mbele M, Hull R, Dlamini Z. African medicinal plants and their derivatives: Current efforts towards potential anti-cancer drugs. *Exp Mol Pathol*. 2017;103(2):121-34. doi: <http://doi.org/10.1016/j.yexmp.2017.08.002>.
 54. Nguyen VT, Sakoff JA, Scarlett CJ. Physicochemical, antioxidant, and cytotoxic properties of *Xao Tam Phan* (*Paramignya trimera*) root extract and its fractions. *Chem Biodivers*. 2017;14(4):1-9. doi: <http://doi.org/10.1002/cbdv.201600396>.
 55. Okubo S, Uto T, Goto A, et al. Berberine Induces Apoptotic Cell Death via Activation of Caspase-3 and -8 in HL-60 Human Leukemia Cells: Nuclear Localization and Structure-Activity Relationships. *Am J Chin Med*. 2017;45(7):1497-511. doi: <http://doi.org/10.1142/S0192415X17500811>.
 56. Pandey S. In vivo antitumor potential of extracts from different parts of *Bauhinia variegata* linn. Against b16f10 melanoma tumour model in c57bl/6 mice. *Appl Cancer Res*. 2017;37:33. doi: <http://doi.org/10.1186/s41241-017-0039-3>.
 57. Rahman MA, Sahabjada, Akhtar J. Evaluation of anticancer activity of *Cordia dichotoma* leaves against a human prostate carcinoma cell line, PC3. *J Tradit Complement Med*. 2017;7(3):315-21. doi: <http://doi.org/10.1016/j.jtcme.2016.11.002>.
 58. Rajavel T, Mohankumar R, Archunan G, et al. Beta sitosterol and Daucosterol (phytosterols identified in *Grewia tiliaefolia*) perturbs cell cycle and induces apoptotic cell death in A549 cells. *Sci Rep*. 2017;7:3418. doi: <http://doi.org/10.1038/s41598-017-03511-4>.
 59. Roman Junior WA, Gomes DB, Zanchet B, et al. Antiproliferative effects of pinostrobin and 5,6-dehydrokavain isolated from leaves of *alpinia*

- zerumbet. *Rev Bras Farmacogn.* 2017;27(5):592-8. doi: <http://doi.org/10.1016/j.bjp.2017.05.007>.
60. Thakor P, Song W, Subramanian RB, et al. Maslinic acid inhibits proliferation of renal cell carcinoma cell lines and suppresses angiogenesis of endothelial cells. *J Kidney Cancer VHL.* 2017;4(1):16-24. doi: <http://doi.org/10.15586/jkcvhl.2017.64>.
 61. Torquato HFV, Goettert MI, Justo GZ, et al. Anti-cancer phytometabolites targeting cancer stem cells. *Curr Genomics.* 2017;18(2):156-74. <http://doi.org/10.2174/1389202917666160803162309>.
 62. Xia X, Cole SPC, Cai T, et al. Effect of traditional Chinese medicine components on multidrug resistance in tumors mediated by P-glycoprotein. *Oncol Lett.* 2017;13(6):3989-96. doi: <http://doi.org/10.3892/ol.2017.5976>.
 63. Zhang Y, Wu W, Kang L, et al. Effect of Aconitum coreanum polysaccharide and its sulphated derivative on the migration of human breast cancer MDA-MB-435s cell. *Int J Biol Macromol.* 2017;103:477-83. doi: <http://doi.org/10.1016/j.ijbiomac.2017.05.084>.
 64. Zhong ZC, Zhao DD, Liu ZD, et al. A new human cancer cell proliferation inhibition sesquiterpene, dryofraterpene a, from medicinal plant *dryopteris fragrans* (L.) Schott. *Molecules.* 2017;22(1):180. doi: <http://doi.org/10.3390/molecules22010180>.
 65. Abu-Darwish MS, Efferth T. Medicinal plants from near east for cancer therapy. *Front Pharmacol.* 2018;9:56. doi: <http://doi.org/10.3389/fphar.2018.00056>.
 66. Al-Dabbagh B, Elhaty IA, Al Hrouf A, et al. Antioxidant and anticancer activities of *Trigonella foenum-graecum*, *Cassia acutifolia* and *Rhazya stricta*. *BMC Complement Altern Med.* 2018;18(1):240. doi: <http://doi.org/10.1186/s12906-018-2285-7>.
 67. Asadi-Samani M, Rafieian-Kopaei M, Lorigooini Z, et al. A screening of growth inhibitory activity of Iranian medicinal plants on prostate cancer cell lines. *Biomedicine (Taipei).* 2018;8(2):8. doi: <http://doi.org/10.1051/bmcdn/2018080208>.
 68. Bonam SR, Wu YS, Tunki L, et al. What has come out from phytomedicines and herbal edibles for the treatment of cancer? *ChemMedChem.* 2018;13(18):1854-72. doi: <http://doi.org/10.1002/cmdc.201800343>.
 69. Chen X, Song L, Hou Y, et al. Reactive oxygen species induced by icaritin promote DNA strand breaks and apoptosis in human cervical cancer cells. *Oncol Rep.* 2018;41(2):765-78. doi: <http://doi.org/10.3892/or.2018.6864>.
 70. Dong R, Chen P, Chen Q. Extract of the medicinal plant pao pereira inhibits pancreatic cancer stem-like cell in vitro and in vivo. *Integr Cancer Ther.* 2018;17(4):1204-15. doi: <http://doi.org/10.1177/1534735418786027>.
 71. Escher GB, Santos JS, Rosso ND, et al. Chemical study, antioxidant, anti-hypertensive, and cytotoxic/cytoprotective activities of *Centaurea cyanus* L. petals aqueous extract. *Food Chem Toxicol.* 2018;118:439-53. doi: <https://doi.org/10.1016/j.fct.2018.05.046>.
 72. Esghaei M, Ghaffari H, Esboei BR, et al. Evaluation of anticancer activity of *Camellia Sinensis* in the Caco-2 colorectal cancer cell line. *Asian Pac J Cancer Prev.* 2018;19(6):1697-701. doi: <http://doi.org/10.22034/APJCP.2018.19.6.1697>.
 73. Fort RS, Barnech JMT, Dourron J, et al. Isolation and structural characterization of bioactive molecules on prostate cancer from mayan traditional medicinal plants. *Pharmaceuticals (Basel).* 2018;11(3):78. doi: <http://doi.org/10.3390/ph11030078>.
 74. Gomes DB, Zanchet B, Locateli G, et al. Antiproliferative potential of solidagenone isolated of *Solidago chilensis*. *Rev Bras Farmacogn.* 2018;28(6):703-9. doi: <https://doi.org/10.1016/j.bjp.2018.09.001>.
 75. Hong J, Fristiohady A, Nguyen CH, et al. Apigenin and luteolin attenuate the breaching of MDA-MB231 breast cancer spheroids through the lymph endothelial barrier in vitro. *Front Pharmacol.* 2018;9:220. doi: <http://doi.org/10.3389/fphar.2018.00220>.
 76. Kuete V, Ngnintedo D, Fotso GW, et al. Cytotoxicity of seputhecarpan D, thonningiol and 12 other phytochemicals from African flora towards human carcinoma cells. *BMC Complement Altern Med.* 2018;18:36. doi: <http://doi.org/10.1186/s12906-018-2109-9>.
 77. Maciel LG, do Carmo MAV, Azevedo L, et al. Hibiscus sabdariffa anthocyanins-rich extract: chemical stability, in vitro antioxidant and antiproliferative activities. *Food Chem Toxicol.* 2018;113:187-97. doi: <https://doi.org/10.1016/j.fct.2018.01.053>.
 78. Malvicini M, Gutierrez-Moraga A, Rodriguez MM, et al. A tricin derivative from *deschampsia antarctica* desv. Inhibits colorectal carcinoma growth and liver metastasis through the induction of a specific immune response. *Mol Cancer Ther.* 2018;17(5):966-76. doi: <http://doi.org/10.1158/1535-7163.MCT-17-0193>.
 79. Ogunlaja OO, Moodley R, Singh M, et al. Cytotoxic activity of the bioactive principles from *Ficus burtt-davyi*. *J Environ Sci Health B.* 2018;53(4):261-75. doi: <https://doi.org/10.1080/03601234.2017.1410385>.
 80. Saeed MEM, Mahmoud N, Sugimoto Y, et al. Betulinic acid exerts cytotoxic activity against multidrug-resistant tumor cells via targeting autocrine motility factor receptor (AMFR). *Front Pharmacol.* 2018;9:481. doi: <http://doi.org/10.3389/fphar.2018.00481>.
 81. Santos JS, Deolindo CTP, Hoffmann JF, et al. Optimized *Camellia sinensis* var. *sinensis*, *Ilex paraguariensis*, and *Aspalathus linearis* blend presents high antioxidant and antiproliferative activities in a beverage model. *Food Chem.* 2018;254:348-58. doi: <https://doi.org/10.1016/j.foodchem.2018.02.021>.

82. Sethi G, Shanmugam MK, Warriar S, et al. Pro-apoptotic and anti-cancer properties of diosgenin: a comprehensive and critical review. *Nutrients*. 2018;10(5):645. doi: <http://doi.org/10.3390/nu10050645>.
83. Sodrul IMD, Wang C, Chen X, et al. Role of ginsenosides in reactive oxygen species-mediated anticancer therapy. *Oncotarget*. 2018;9(2):2931-50. doi: <http://doi.org/10.18632/oncotarget.23407>.
84. Soyngbe OS, Mongalo NI, Makhafola TJ. In vitro antibacterial and cytotoxic activity of leaf extracts of *Centella asiatica* (L.) Urb, *Warburgia salutaris* (Bertol. F.) Chiov and *Curtisia dentata* (Burm. F.) C.A.Sm - medicinal plants used in South Africa. *BMC Complement Altern Med*. 2018;18:315. doi: <http://doi.org/10.1186/s12906-018-2378-3>.
85. Tayeh Z, Ofir R. *Asteriscus graveolens* extract in combination with cisplatin/etoposide/doxorubicin suppresses lymphoma cell growth through induction of caspase-3 dependent apoptosis. *Int J Mol Sci*. 2018;19(8):2219. doi: <http://doi.org/10.3390/ijms19082219>.
86. Uen WC, Lee BH, Shi YC, et al. Inhibition of aqueous extracts of *Solanum nigrum* (AESN) on oral cancer through regulation of mitochondrial fission. *J Tradit Complement Med*. 2018;8(1):220-5. doi: <https://doi.org/10.1016/j.jtcme.2017.05.011>.
87. Zhang T, Wang L, Duan DH, et al. Cytotoxicity-guided isolation of two new phenolic derivatives from *Dryopteris fragrans* (L.) Schott. *Molecules*. 2018;23(7):1652. doi: <http://doi.org/10.3390/molecules23071652>.
88. Asl EA, Mehrabadi JF, Afshar D, et al. Apoptotic effects of *Linum album* extracts on AGS human gastric adenocarcinoma cells and ZNF703 oncogene expression. *Asian Pac J Cancer Prev*. 2018;19(10):2911-6. doi: <http://doi.org/10.22034/APJCP.2018.19.10.2911>.
89. Aumeeruddy MZ, Mahomoodally MF. Combating breast cancer using combination therapy with 3 phytochemicals: Piperine, sulforaphane, and thymoquinone. *Cancer*. 2019;125(10):1600-11. doi: <http://doi.org/10.1002/cncr.32022>.
90. Cordeiro YG, Rochetti AL, Souza VC, et al. Antineoplastic effect of procyanidin-rich extract of *Lafoensia Pacari* in lung carcinoma cells. *Braz Arch Biol Technol*. 2019;62:e19160638. doi: <http://doi.org/10.1590/1678-4324-2019160638>.
91. De D, Chowdhury P, Panda SK, et al. Ethanolic extract of leaf of *Dillenia pentagyna* reduces in-vitro cell migration and induces intrinsic pathway of apoptosis via downregulation of NF- κ B in human NSCLC A549 cells. *J Cell Biochem*. 2019;120(12):19841-57. doi: <http://doi.org/10.1002/jcb.29289>.
92. Iqbal J, Abbasi BA, Ahmad R, et al. Potential phytochemicals in the fight against skin cancer: current landscape and future perspectives. *Biomed Pharmacother*. 2019;109:1381-93. doi: <https://doi.org/10.1016/j.biopha.2018.10.107>.
93. Lim CP, Yam MF, Asmawi MZ, et al. Cytostatic and antiproliferative activities of f5 fraction of crinum amabile leaf chloroform extract showed its potential as cancer chemotherapeutic agent. *Evid Based Complement Alternat Med*. 2019;2019:7521504. doi: <http://doi.org/10.1155/2019/7521504>.
94. Mfengwana PH, Mashele SS, Manduna IT. Cytotoxicity and cell cycle analysis of *Asparagus larinicus* Burch. and *Senecio asperulus* DC. on breast and prostate cancer cell lines. *Heliyon*. 2019;5(5):e01666. doi: <https://doi.org/10.1016/j.heliyon.2019.e01666>.
95. Sinha K, Chowdhury S, Banerjee S, et al. Lupeol alters viability of SK-RC-45 (Renal cell carcinoma cell line) by modulating its mitochondrial dynamics. *Heliyon*. 2019;5(8):e02107. doi: <https://doi.org/10.1016/j.heliyon.2019.e02107>.
96. Wang N, Fan Y, Yuan CM, et al. Selective ERK1/2 agonists isolated from *Melia azedarach* with potent anti-leukemic activity. *BMC Cancer*. 2019;19:764. doi: <http://doi.org/10.1186/s12885-019-5914-8>.
97. Zahra MH, Salem TAR, El-Aarag B, et al. *Alpinia zerumbet* (Pers.): Food and medicinal plant with potential in vitro and in vivo anti-cancer activities. *Molecules*. 2019;24(13):2495. doi: <http://doi.org/10.3390/molecules24132495>.

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